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Targeting a critical step in fungal hexosamine biosynthesis

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Edited by Gerald W. Hart

Aspergillus fumigatus is a human opportunistic fungal pathogen whose cell wall protects it from the extracellular environment including host defenses. Chitin, an essential component of the fungal cell wall, is synthesized from UDP-GlcNAc produced in the hexosamine biosynthetic pathway. As this pathway is critical for fungal cell wall integrity, the hexosamine biosynthesis enzymes represent potential targets of antifungal drugs. Here, we provide genetic and chemical evidence that glucosamine 6-phosphate *N*-acetyltransferase (Gna1), a key enzyme in this pathway, is an exploitable antifungal drug target. *GNA1* deletion resulted in loss of fungal viability and disruption of the cell wall, phenotypes that could be rescued by exogenous GlcNAc, the product of the Gna1 enzyme. In a murine model of aspergillosis, the Δ *gna1* mutant strain exhibited attenuated virulence. Using a fragment-based approach, we discovered a small heterocyclic scaffold that binds proximal to the Gna1 active site and can be optimized to a selective submicromolar binder. Taken together, we have provided genetic, structural, and chemical evidence that Gna1 is an antifungal target in *A. fumigatus*.

Aspergillus spp., *Candida* spp., and *Cryptococcus* spp. are opportunistic human pathogens that together account for most global invasive fungal infections. Considered a significant and growing challenge to human health (1, 2), infections attributable to *Aspergillus fumigatus* are a significant cause of morbidity and mortality in an ever-expanding group of patients. For patients undergoing hematopoietic stem cell transplantation and in particular allogeneic grafts, invasive aspergillosis is an important cause of pulmonary-related mortality (3). Chronic and allergic forms are rarely life threatening, however, they are estimated to have global burdens of ~1.2 million and 4.8 million people, respectively (4, 5). Moreover, they may affect patients with an intact immune system but have other medical co-morbidities, placing them at a higher risk of fungal sensitization and subsequent allergic disease. A recent phenomenon in both immunosuppressed and immunocompetent individuals is post-influenza aspergillosis in the intensive care unit (6).

As both fungi and mammals are eukaryotes, antifungal drug development faces the fundamental challenge of selective toxicity. Structures that are unique to fungi or that bear exploitable dissimilarities, such as the cell membrane and the cell wall, are targeted by the polyenes (7), azoles (8), and echinocandins (9), respectively. Together with flucytosine (10), they represent the only antifungal classes clinically licensed for the prevention and treatment of aspergillosis (3) and offer limited degrees of effectiveness through a restricted spectrum of cellular targets, poor clinical and physicochemical properties, and rising rates of drug resistance (11), particularly exacerbated through extensive agricultural overuse of antifungal azoles (2, 12). A lack of innovation, investment, and infrastructure in pharmaceutical antifungal research has been cited for the dearth of preclinical candidates despite a global market worth in excess of U.S. \$6 billion (13). Traditionally, antifungal agents were discovered by screening large libraries of natural products or synthetic small molecules for their fungistatic and, preferably, fungicidal properties (14). The polyenes and echinocandins were both derived from natural product screening. Unfortunately, many subsequent screens rediscovered the same basic chemical scaffolds (14). The antifungal pipeline has displayed modest progression (15), with only a single new antifungal class currently undergoing late-stage clinical trials (Phase IIb, NCT03583164; RRID: SCR_002309). Although the introduction of additional azoles such as isavuconazole (16, 17) provides greater options to reduce side effects and simplify dosing regimens, the development of resistance is not adequately addressed by new members of this widely used class (18).

A major characteristic feature of fungi is the presence of a highly ordered carbohydrate cell wall that provides protection against chemical and mechanical stresses. The cell wall is a dynamic, interlaced, and only partially defined polysaccharide structure that is essential for survival (19) and is considered an attractive source of novel antifungal targets because of its absence in humans. The *A. fumigatus* cell wall predominantly consists of a central core of fibrils composed of branched β -1,3 glucan crosslinked to chitin. Polysaccharides such as β -1,3–1,4-glucan and galactomannan are covalently bound to this complex, whereas those contained in the surrounding cement include α -1–3 glucan. Cell wall biosynthesis and remodeling is a highly organized and complex process, although many of the key steps remain to be fully elucidated (19). Polysaccharide synthesis of β -1,3 glucan and chitin occur at the fungal cell membrane using intracellular sugar nucleotide donors as substrates.

This article contains supporting information.

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Chitin is an integral structural component of the cell wall in *A. fumigatus*, contributing to rigidity, and consists of a linear polymer of $\beta(1-4)$ -linked *N*-acetyl-D-glucosamine (GlcNAc). Chitin synthesis remains an attractive drug target; however, targeting the entire chitin synthase family is not a viable option because of predicted differences in structure and the membrane-bound location of the eight proteins (20).

An alternative target to chitin synthases is biosynthesis of the sugar nucleotide substrate UDP-GlcNAc that is also utilized for the synthesis of GPI anchors and *N*- and *O*-linked glycans (21). Four highly conserved enzymes in UDP-GlcNAc biosynthesis form the hexosamine biosynthetic pathway (22), which is believed to represent the only endogenous source of UDP-GlcNAc within fungal cells (23). Approximately 2–5% of glucose that enters the cell is directed into this pathway where it is phosphorylated to glucose-6-phosphate (Glc-6P) and subsequently converted to fructose-6-phosphate (Fru-6P). Four enzymes catalyze the remaining steps of this pathway. Glucosamine-6-phosphate synthase (Gfa1) converts Fru-6P to glucosamine-6-phosphate (GlcN-6P). This undergoes acetylation by glucosamine-6-phosphate *N*-acetyltransferase (Gna1) to GlcNAc-6-phosphate (GlcNAc-6P). The next step involves isomerization by phospho-acetyl-glucosamine mutase (Agm1) to yield GlcNAc-1-phosphate (GlcNAc-1P) before uridylation by GlcNAc-1P-uridylyltransferase (Uap1) to produce UDP-GlcNAc. This pathway is highly regulated in filamentous fungi, yeast, and higher eukaryotes (23). Three of the enzymes (Gfa1, Agm1, and Uap1) have been genetically validated in *A. fumigatus* as essential for growth under *in vitro* laboratory conditions (24–26).

Gna1 (EC 2.3.1.4) is a member of the Gcn5-related *N*-acetyltransferase (GNAT) superfamily and is present in yeast such as *Candida* spp. (e.g. *Candida albicans*, *Candida glabrata*, and *Candida auris*) and *Saccharomyces cerevisiae*. Genetic studies in *S. cerevisiae* have demonstrated that deletion of *GNA1* is lethal (27). In *C. albicans* *GNA1* is essential for *in vitro* viability and attenuated virulence in a murine model of disseminated candidiasis (28, 29). Inactivation of the murine *GNA1* homologue is lethal as it is implicated in mitotic membrane fusion events and this further reinforces a critical role for this enzyme in higher eukaryotes (30). Although evolutionarily and structurally distinct from higher eukaryotic organisms, the *GNA1* orthologues recently identified in parasitic protozoan apicomplexans have been suggested as potential therapeutic targets (31).

To date, Gna1 in *A. fumigatus* is unexplored as an antifungal drug target using a combination of genetic and chemical methods. Inhibitors that chemically phenocopy a *GNA1* deletion have not been reported in any organism, and the high degree of sequence homology between eukaryotic *GNA1* family members likely precludes fungal selectivity. Here, we report a multidisciplinary approach providing genetic, chemical, and structural evidence supporting *A. fumigatus* Gna1 as an antifungal target and describe a fragment that exploits a previously unidentified pocket selective for the fungal enzyme.

Results

Disruption of *gna1* leads to a terminal phenotype that is rescued by exogenous GlcNAc

To investigate the requirement for Gna1 in *A. fumigatus* UDP-GlcNAc biosynthesis, we deleted *GNA1* by homologous recombination. A $\Delta gna1$ strain was constructed by replacing the coding sequence with a *pyrG* selection marker (Fig. S1). Transformants were evaluated by phenotypic and PCR screening before verification by Southern blotting (Fig. S1). Using a similar approach, *GNA1* (with an N-terminal His tag) was re-introduced into the $\Delta gna1$ mutant (Fig. S1). Although suitable for initial *in vitro* studies, it was imperative that the reconstituted strain restored *pyrG* prototrophy as *pyrG*– *A. fumigatus* strains are highly attenuated in murine models of aspergillosis (32). To address this, the *pyrG*– strain was targeted with random integration of *A. fumigatus pyrG* into the chromosome to generate a *pyrG*+ reconstituted strain (Fig. S1).

In vitro growth of *A. fumigatus* $\Delta gna1$ was completely inhibited in glucose (Glc)-supplemented media and restored to comparable levels upon replacement of glucose with GlcNAc, the product of the Gna1 enzyme (Fig. 1A). This demonstrates that *GNA1* is essential for viability of *A. fumigatus* in standard *in vitro* laboratory media rich in glucose and concurs with previous work in yeast (27, 29). We considered the physiological relevance of these Glc and GlcNAc concentrations because failure to correlate with physiological nutritional availability may incorrectly assign a gene as essential for growth. The glucose concentration used is 20-fold greater than the mean physiological human blood glucose level (110 mM versus ~5.5 mM) and in lung alveoli glucose concentrations are further reduced (~0.1 mM) (33). Specific human physiological levels of GlcNAc are unspecified in the literature but regarded as low and subject to fluctuation. We initially investigated rescue of $\Delta gna1$ on solid media supplemented with Glc (0.1 mM) and a simulated range of physiological GlcNAc concentrations, above and below previously reported levels (29) (Fig. 1A). The $\Delta gna1$ growth phenotype in the presence of low GlcNAc levels (1.5 μ M) was characterized by a complete loss of hyphae and melanin pigment. High GlcNAc levels (150 μ M, 100-fold increase) partially restored hyphal development, but colonies were still strikingly devoid of pigmentation and overall radial growth was less than the parental and reconstituted strain. We describe these characteristics as a terminal phenotype.

To dissect the interplay between exogenous Glc and GlcNAc, we developed a checkerboard assay to assess 285 different nutritional combinations of these to explore possible concentrations encountered by *A. fumigatus* in the human host environment (Fig. 1B). Heat map analysis demonstrated that loss of *GNA1* is associated with a terminal phenotype defective in conidial pigmentation and hyphal growth across a range of estimated physiological concentrations.

The observed $\Delta gna1$ terminal phenotype is unlikely to be a result of a Gna1 scaffolding effect as metabolic supplementation with GlcNAc is restorative, indicating that disruption of Gna1 enzymatic function is sufficient to elicit the observed reduction in fungal viability. Taken together, these experiments

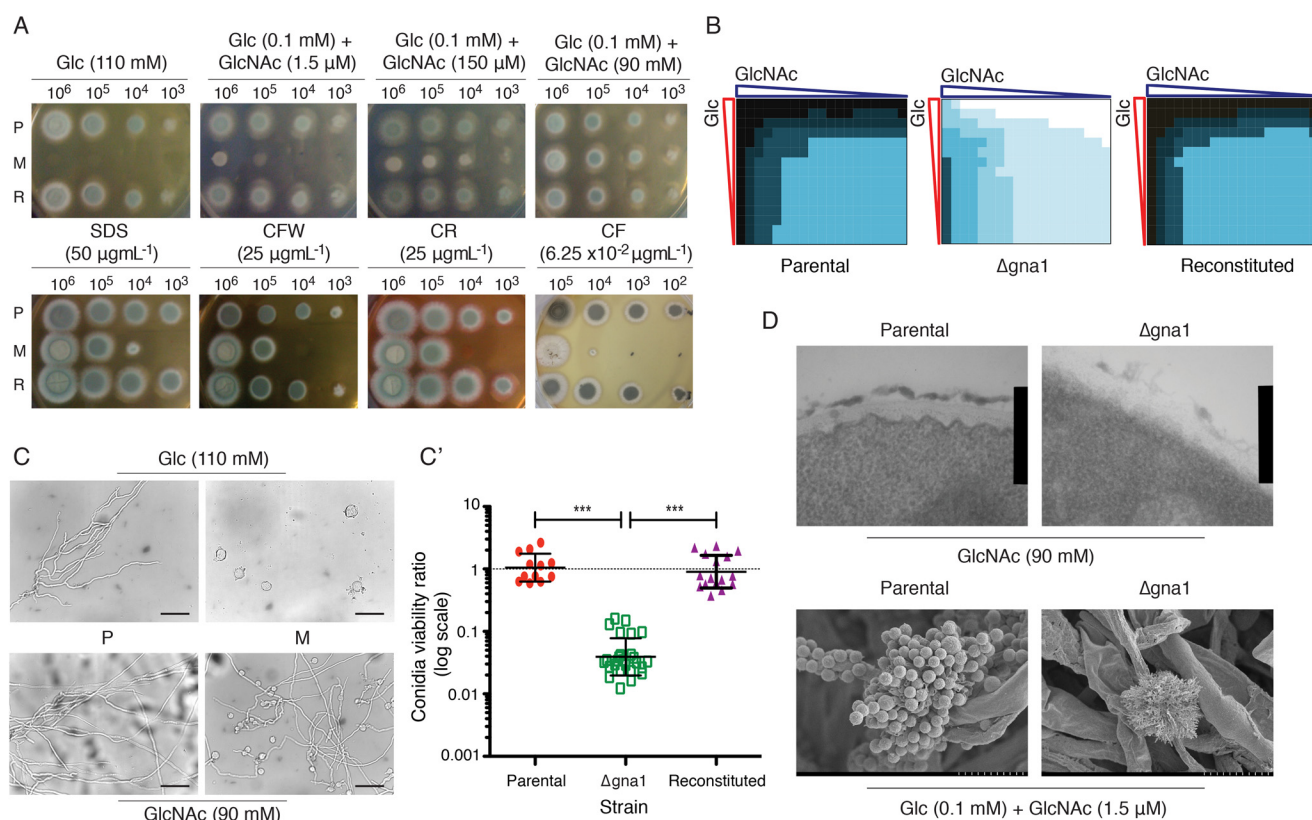


Figure 1. Disruption of *gna1* leads to a terminal cell wall phenotype partially rescued by GlcNAc. A, upper panel, colony morphology after 48 h incubation following inoculation of viable *A. fumigatus* conidia on CM supplemented with Glc and GlcNAc (as indicated). The morphology of $\Delta gna1$ in the central two images is indicative of the terminal phenotype. Lower panel, serial dilutions of viable conidia were inoculated onto solid CM (0.1 mM Glc + ≥ 50 mM GlcNAc) supplemented with SDS, Calcofluor White (CFW), Congo Red (CR), or caspofungin (CF) and incubated for 48 h. Images are representative of results obtained from three independent experiments (n = 3). B, heat map of growth after 48 h incubation (inoculum 1×10^3 viable conidia) in a checkerboard assay containing combinations of Glc and GlcNAc from 0 to 50 mM (2-fold serial dilutions). The parental, $\Delta gna1$ mutant, and reconstituted strain were scored numerically from 1 (no growth/inoculum spot) to 7 (confluent growth with dark pigmented conidia). Each square denotes an individual condition and represents the mean growth from three independent experiments (n = 3). Black (maximum growth) to white (no growth). C, exogenous Glc arrests conidial germination and GlcNAc rescue fails to fully restore viability of $\Delta gna1$. Left panel, conidia were inoculated into liquid CM containing Glc (upper) and GlcNAc (lower), incubated for up to 24 h and examined using a Leica light microscope at $\times 40$ magnification. Scale bar: 0.05 mm. C', ratio of viable conidia versus total number of conidia. Each data point represents an individual replicate from a minimum of three independent experiments; horizontal lines represent the mean \pm S.D. (n = 3). A ratio of 1 denotes no difference. ***, $p < 0.0001$. D, viable conidia (1×10^5) were inoculated onto solid CM supplemented with Glc and GlcNAc and incubated for 16 h (hyphae, upper panel) or 96 h (conidia, lower panel). Samples were fixed, processed, and examined using EM (upper panel, TEM; lower panel, SEM). Scale bar: upper panel, 361 nm; lower panel, 10 μ m. P, parental; M, $\Delta gna1$ mutant; R, reconstituted *A. fumigatus* strains. All incubations were at 37 $^{\circ}$ C.

show that disruption of *gna1* leads to a terminal phenotype that is rescued by exogenous GlcNAc.

The $\Delta gna1$ strain displays a cell wall phenotype

Next, we investigated whether rescue of *GNA1* with exogenous GlcNAc induces a cell wall stress response. *A. fumigatus* harbors cell wall integrity pathways that are activated under conditions of stress by signaling pathways such as the mitogen activated protein kinase cascades (34). As the terminal phenotype did not yield sufficient growth for comparative analyses, this experiment was performed using a GlcNAc concentration of 90 mM to rescue growth. At concentrations of $\leq 10^4$ viable conidia, the $\Delta gna1$ mutant displayed increased sensitivity to agents compromising the cell wall and membrane, suggesting the presence of cell wall defects and/or loss of membrane integrity (Fig. 1A). Calcofluor White interacts with chitin, leading to disruption of chitin polymer assembly and impairing the corresponding linkages to other cell wall components (35). Increased susceptibility to Congo Red suggests a reduction of β -1,3 and β -1,6 glucans. Increased sensitivity to sodium dodecyl sulfate

(SDS) reflects a compromised membrane that may be because of differences in the chitin layer. Finally, we investigated whether loss of *A. fumigatus* *GNA1* enhanced the effects glucan synthesis inhibition by the echinocandin caspofungin (Fig. 1A). Exposure to caspofungin induced a terminal phenotype in the $\Delta gna1$ mutant ($\geq 10^4$ conidia) and was fungicidal at lower concentrations. This provides evidence of synergistic *in vitro* activity of a β -glucan synthase inhibitor together with the $\Delta gna1$ mutant.

To probe the effects of Glc on germination, parental and $\Delta gna1$ mutant germlings were examined by light microscopy (Fig. 1C). Under conditions tested the $\Delta gna1$ mutant was non-viable (Fig. 1A) and examination of the conidia demonstrated a subset that was enlarged and highly disorganized with evidence of multiple, rudimentary germ-tubes whose development was prematurely arrested (Fig. 1C). Despite macroscopically comparable phenotypes, the $\Delta gna1$ mutant conidia incubated in the presence of 90 mM GlcNAc also displayed abnormalities compared with the parental strain (Fig. 1C). Although hyphae were evident at 24 h, there was also evidence of conidial

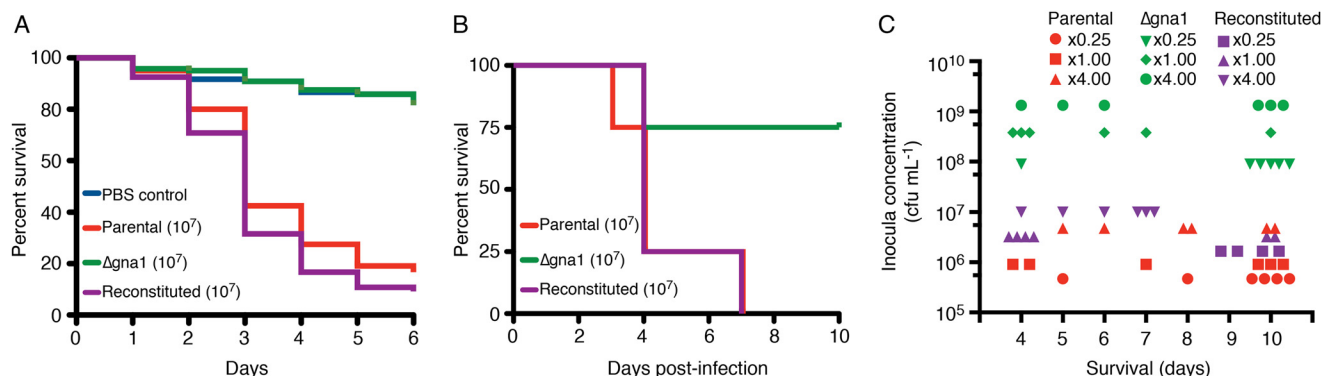


Figure 2. *GNA1* contributes to *A. fumigatus* pathogenicity in invertebrate and murine models of aspergillosis. A, *G. mellonella* were inoculated with 5×10^5 conidia of *A. fumigatus* (parental, $\Delta gna1$ mutant, or reconstituted strains) or PBS as a vehicle control. Survival was monitored daily as evident by movement and lack of melanization. Data are combined from four independent experiments ($n = 4$ with a combined total of 120 larvae per group). $p < 0.0001$. B, neutropenic male CD1 mice were exposed to 1×10^7 *A. fumigatus* aerosolized conidia from the parental, $\Delta gna1$ mutant, and reconstituted strains in an inhalational chamber ($n = 4$ per group). C, neutropenic male CD1 mice were exposed to a trio of refined doses of *A. fumigatus* aerosolized conidia from the parental, $\Delta gna1$ mutant, and reconstituted strains in an inhalational chamber ($n = 6$ per group). A reference dose of $\times 1.00$ was set to that observed to induce signs of disease from day 3. Each symbol represents an individual animal. In B and C, mice were monitored for up to 10 days post infection and any mouse showing clinical signs of disease or distress was culled immediately and death being recorded as the next day.

enlargement and a lack of polarized hyphal growth with some $\Delta gna1$ germ tubes unable to branch (Fig. 1C). We hypothesize that this subpopulation of abnormal conidia accounts for the significant reduction in viability of the *A. fumigatus* $\Delta gna1$ strain (Fig. 1C'). GlcNAc is unable to completely rescue this phenotype. This suggests a possible cell wall phenotype because of reduced flux through the hexosamine biosynthetic pathway.

Defects in the cell surface architecture of the parental and $\Delta gna1$ mutant strains were also revealed by both scanning electron microscopy (SEM) and transmission electron microscopy (TEM). SEM analysis demonstrated that the $\Delta gna1$ mutant grown under terminal phenotype conditions (0.1 mM Glc and 1.5 μ M GlcNAc) displayed conidiophore collapse (Fig. 1D). The vesicular head was grossly abnormal with the presence of “naked” phialides and no obvious conidia compared with the parental strain (Fig. 1D). TEM analysis revealed a disorganized cell wall ultrastructure for the $\Delta gna1$ mutant, with a less-defined chitin layer and a more dispersed outer mannoprotein layer (Fig. 1D). These data suggest that loss of *GNA1* is associated with a defective and potentially more porous cell wall and provides a molecular basis for the observed synergy with caspofungin. Taken together, these data show that the $\Delta gna1$ strain displays a cell wall phenotype.

GNA1 contributes to *A. fumigatus* pathogenicity in models of aspergillosis

Ascertaining the physiological implications of *GNA1* loss by translating *in vitro* phenotypic characterizations into *in vivo* infection models is an important step in providing genetic validation as an antifungal target. In particular, it is critical to determine whether *in vivo* GlcNAc scavenging systems could compensate for *GNA1* loss. Nonvertebrate mini-host model systems such as the greater wax moth larvae (*Galleria mellonella*) represent a rapid initial screening strategy (36). Larval inoculation resulted in a mean survival time of 3 days for the parental and reconstituted strains whereas the $\Delta gna1$ mutant was avirulent with a survival profile indistinguishable from the PBS control group (Fig. 2A).

A disadvantage of *G. mellonella* is that it fails to represent the natural inhalation route of *Aspergillus*. To recapitulate inhalational acquisition of *A. fumigatus* and provide insight into the physiological interplay of alveolar glucose and GlcNAc levels in circumventing *GNA1* loss, we used an established neutropenic murine model of invasive pulmonary aspergillosis (37). A preliminary study was performed using three concentrations of viable *A. fumigatus* conidia and the contribution of the $\Delta gna1$ mutant to pathogenicity was compared with the parental and reconstituted strains. In this experiment, the $\Delta gna1$ mutant was less virulent (Fig. 2B). The median survival time was 4 days for the parental and reconstituted strains whereas a value could not be determined for the $\Delta gna1$ mutant (75% survival). This finding was confirmed in a subsequent study (Fig. 2C) using doses refined for each strain designed to induce signs of invasive aspergillosis after ~3 days post inhalation (see supporting information). Although the mean survival times between *A. fumigatus* strains are not directly comparable because of differences in dose, there was an 83% survival rate to day 10 for mice exposed to the lowest concentration ($\times 0.25$) of the $\Delta gna1$ mutant strain compared with 33% and 0% for the parental and reconstituted strains at their highest concentrations ($\times 4.00$) (see Table S2). Approximately 1 to 2 log higher concentrations of $\Delta gna1$ were required to induce clinical disease (Fig. 2C). Overall these findings suggest that a degree of metabolic (GlcNAc) rescue of $\Delta gna1$ may occur *in vivo*. Nevertheless, these data show that *GNA1* contributes to *A. fumigatus* pathogenesis in models of aspergillosis.

Discovery of small molecules that target the *Gna1* dimer interface

There are currently no selective small molecule inhibitors for the *Gna1* class of enzymes to attempt to chemically phenocopy the genetic phenotype. Mechanistically, *A. fumigatus* (Af) *Gna1* facilitates *N*-acetylation of GlcN-6P by promoting direct nucleophilic attack at the thioester carbonyl group of the acetyl coenzyme A (AcCoA) cofactor by the sugar-phosphate amino group (38). Although sharing only 30% sequence identity,

Targeting a critical step in fungal hexosamine biosynthesis

structurally *AfGna1* and *Homo sapiens* (*Hs*) Gna1 adopt the same overall fold and domain architecture, with highly conserved regions localized to the AcCoA-binding site with minor differences in the sugar substrate-binding region. These differences are localized to amino acids involved in sugar-phosphate group recognition, which in the fungal enzyme utilize side chain functionality that is less polar in character (38, 39). In an attempt to circumvent issues of high structural homology between human and fungal Gna1, we deployed a fragment-based approach (40) to discover small molecules that target previously unidentified pockets that are selective for the fungal enzyme. Bio-layer interferometry (BLI) was used to identify Gna1 binders from a 650-compound in-house fragment library (41–43). This produced an initial hit rate of 5.7% (Fig. S2). The top hit (Fig. 3A, 1) displayed stoichiometric binding and high ligand efficiency ($LE = 0.57 \text{ kcal mol}^{-1} \text{ NHA}^{-1}$ for 1), with a $6 \mu\text{M}$ equilibrium dissociation constant which was confirmed by isothermal titration calorimetry (Fig. S2).

We next used X-ray crystallography to determine the binding mode of 1. *AfGna1* was co-crystallized with AcCoA and soaked into solutions of 1. Synchrotron diffraction data to 1.6 \AA (Fig. 3, Table 1) were collected and used to solve the structure by molecular replacement. Previous work has shown that *A. fumigatus* Gna1 possesses a classic GNAT α/β -fold, forming an obligate homodimer. This dimer shows extensive secondary structure swapping with substrate and cofactor binding to the two active sites that are formed by channels on the surface of the enzyme, composed of amino acids from both *AfGna1* monomers (39). Intriguingly, electron density corresponding to fragment hit 1 was observed in a pocket buried at the dimer interface in the core of the enzyme, binding across a crystallographic 2-fold axis that generates the *AfGna1* homodimer and the symmetry-related molecule of fragment hit 1, visible in the *AfGna1*-1 co-complex structure (Fig. 3B). This hydrophobic pocket ($\sim 400 \text{ \AA}^3$) accommodates fragment hit 1, which is positioned $\sim 8 \text{ \AA}$ from both GlcN-6P substrate-binding sites with a histidine residue (His-127) from each monomer trapping the fragment in a π - π stacking arrangement (Fig. 3B). Critically, *AfGna1* His-127 is substituted with an arginine (Arg-118) in the human enzyme, blocking the pocket and importantly, preventing fragment binding to *HsGna1* (Fig. 3B, Fig. S3), whereas His-127 is conserved among clinically relevant fungal species (Fig. S3). With the exception of Arg-118, the amino acids lining the fragment-binding pocket are conserved between clinically relevant fungal Gna1 orthologues and *HsGna1* (Fig. S3).

To develop the fragment scaffold toward *AfGna1*-selective inhibition, the binding characteristics of the fragment within the dimer interface pocket were explored to identify the structural requirements for optimal *AfGna1* binding (Fig. S2). The staggered bi-ring system of 1 in the *AfGna1*-1 complex is likely a consequence of reducing steric repulsion between the *ortho*-chloro substituent of the aryl ring system and the adjacent 1,2,4-triazole (dihedral angle, θ ; $\sim 51^\circ$). This results in optimal positioning of 1 to facilitate the important π - π stacking interactions with the His-127 residues present in the pocket (Fig. 3B). It appeared essential to maintain this bi-ring arrangement as loss or replacement of the *ortho*-substituent increased or ablated binding affinity (Fig. S2).

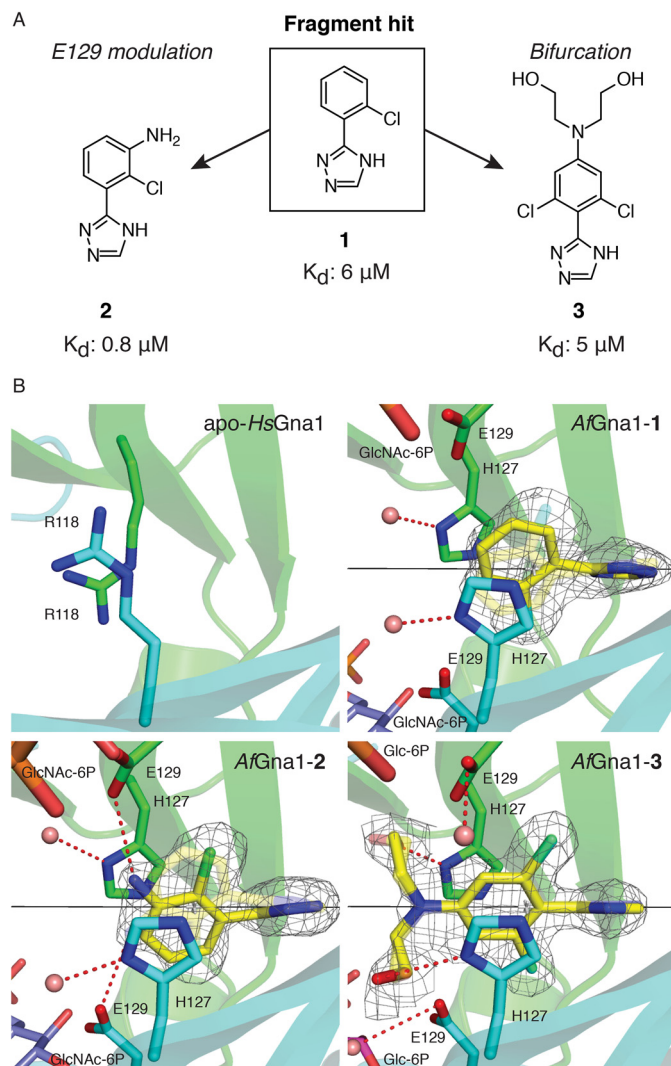


Figure 3. Derivatives of a fragment screen hit selectively target an unusual Gna1 pocket close to the active site. A, chemical structures and binding affinities of fragment-hit (1) and derivatives (2 and 3). B, panels showing the structures of apo-*HsGna1* (PDB ID 2HUZ (91)) and co-complex structures of *AfGna1*-1, *AfGna1*-2, and *AfGna1*-3. Gna1 monomer units (cyan and green); bound small molecules and crystallographic symmetry-related molecules (yellow and transparent sticks, respectively); GlcNAc-6P (blue sticks); Glc-6P (plum sticks); water molecules (pink spheres); hydrogen-bond interactions (red dashed lines); crystallographic axis (black lines) are shown. In all co-complex structures, small molecule electron density was observed binding across the crystallographic 2-fold axis that generates the *AfGna1* homodimer. The symmetry-equivalent molecules of fragment hit 1, 2, and 3 are shown as transparent sticks. In apo-*HsGna1*, the novel dimer interface-binding pocket is occluded by Arg-118 residues preventing ligand binding. In *AfGna1*, substitution of Arg-118 by His-127 facilitates π - π stacking with 1, 2, and 3 forming the basis of fungal selective fragment binding. Superposition of enzymatic product, GlcNAc-6P (PDB ID 2VXK (89)) onto co-complex structures *AfGna1*-1 and *AfGna1*-2 demonstrates the proximity of the fragment hit and derivatives to the Glu-129 side chains and substrate binding sites. The AcCoA cofactor is not visible from selected viewpoint. The $F_o - F_c$ map for the bound small molecules (fragment hit 1, 2, and 3) are shown as a mesh (gray) contoured to 2.5σ .

With the *ortho*-chloro substituted scaffold identified, we considered the close proximity of the glutamic acid residue (Glu-129) that separates fragment hit 1 and the substrate-binding site from each respective *AfGna1* monomer. Superposition of *AfGna1*-GlcNAc-6P complex (the *N*-acetylated enzymatic product of GlcN-6P; PDB ID 2VXK (89)) onto the co-complex structure of *AfGna1* with 1 illustrates the close proximity of the

Table 1**Crystallographic data collection and refinement statistics**

Statistics for the highest-resolution shell are shown in parentheses.

| | <i>AfGna1-1-AcCoA</i> | <i>AfGna1-2-AcCoA</i> | <i>AfGna1-3-AcCoA-(Glc-6P)</i> |
|---|-----------------------|-----------------------|--------------------------------|
| Beamline | <i>DLS-IO4-1</i> | <i>ESRF-ID30A-3</i> | <i>ESRF-ID30A-1</i> |
| Resolution (Å) | 56.46 (1.64–1.58) | 40.32 (1.80–1.74) | 58.2 (2.08–2.01) |
| Space group | C 2 2 21 | C 2 2 21 | C 2 2 21 |
| Unit cell (Å) <i>a</i> , <i>b</i> , <i>c</i> | 67.64, 102.51, 56.62 | 70.21, 101.2, 56.37 | 70.96, 101.71, 55.91 |
| $\alpha = \beta = \gamma$ (°) | 90 | 90 | 90 |
| Total reflections | 150,350 (5090) | 94,486 (4877) | 53,191 (2813) |
| Unique reflections | 26,623 (2681) | 20,528 (2069) | 13,064 (1243) |
| Multiplicity | 5.6 (3.8) | 4.6 (4.7) | 4.0 (4.2) |
| Completeness (%) | 97 | 98 | 94.6 |
| <i>I</i> / σ (<i>I</i>) | 11.8 (0.7) | 16.2 (2.3) | 14.5 (2.4) |
| <i>R</i> _{merge} (%) | 6.2 (123) | 4.1 (49.0) | 4.9 (57.8) |
| <i>CC</i> _{1/2} | 99.8 | 99.9 | 99.9 |
| <i>R</i> _{work} / <i>R</i> _{free} (%) | 20.0/22.7 | 19.0/23.5 | 21.1/26.4 |
| Average B-factor (Å ²) | 38.0 | 35.7 | 46.6 |
| Protein | 37.90 | 35.48 | 46.25 |
| Ligand | 32.77 | 29.38 | 47.62 |
| Solvent | 47.42 | 42.93 | 54.03 |
| No. of residues | 164 | 165 | 165 |
| No. of water molecules | 108 | 88 | 45 |
| Root mean square deviation | | | |
| Bonds (Å) | 0.012 | 0.013 | 0.009 |
| Angles (°) | 1.89 | 1.89 | 1.81 |
| PDB code | 6TDH | 6TDG | 6TDF |

Glu-129 side chains to the fragment hit 1 (3.4 Å) and to the substrate-binding sites (8.2 Å) (Fig. 3B). Previous Gna1 studies have indicated that orthologous glutamate residues, although not playing a catalytic role, contribute to substrate and cofactor binding through direct contacts and through hydrogen bond-mediated water interactions (27, 44, 45). Indeed, our data demonstrate that mutations which remove the Glu-129 carboxylate (E129A) or substitute it with the corresponding amide (E129Q) reduced enzymatic activity compared with WT *AfGna1* (Fig. S4). As such, we hypothesized that the close proximity of Glu-129 residues to fragment hit 1 may offer scope to develop derivatives that exploit the Glu-129 side-chain functionality to both improve fragment-binding affinity and elicit enzyme inhibition through perturbation of Glu-129-mediated interactions (Fig. 3A, *E129 modulation approach*).

To explore the feasibility of harnessing Glu-129 fragment proximity toward enzyme binding and inhibition, a *meta*-anilino group was installed into the scaffold. This resulted in fragment derivative 2, which displayed submicromolar affinity (0.8 μM, Fig. 3A). Derivative 2 also displayed improved ligand efficiency compared with fragment hit 1 as a result of this minimal scaffold addition, resulting in a highly efficient binder (LE = 0.64 kcal mol^{−1} NHA^{−1} for 2). Overall, 2 maintains the important π - π binding interaction observed in the *AfGna1-1* complex structure. As observed in the *AfGna1-2* complex structure, the introduction of the *meta*-anilino group causes a positional shift of 2 (32° from fragment hit 1), facilitating a hydrogen bond interaction between 2 and the Glu-129 side chain that likely drives the observed enhancement in binding affinity (Fig. 3). Although derivative 2 did not show inhibition against *AfGna1* in a recombinant enzyme assay, exploiting the proximity of the Glu-129 side chains may be a route to improving binding potency, through provision of an anchoring point from which to introduce chemical functionality to modulate the position or protonation state of the Glu-129 residues to achieve inhibition.

As an alternative approach to achieving *AfGna1* inhibition, we aimed to exploit the protein tunnels that connect the dimer interface fragment binding pocket to the substrate-binding sites (distance of 1 to substrate-binding site = ~8 Å, Fig. 3). Extension of the fragment scaffold into the active sites by traversing these tunnels would effectively generate a competitive enzyme inhibitor by blocking substrate binding at both *AfGna1* active sites (Fig. 3A, *bifurcation approach*).

In contrast to Glu-129 modulation, the success of bifurcation is predicated upon the ability of fragment hit 1 derivatives to both successfully reach the buried dimer interface binding pocket and be spatially accommodated despite the concomitant increase in fragment molecular size, which could conceivably hinder binding on both accounts. To test the ability of a bifurcated derivative of 1 to bind *AfGna1*, 3 was synthesized. This bifurcated fragment introduces an *N,N*-diethyl alcohol moiety into the scaffold and displays a binding affinity of the same order as fragment hit 1 (5 μM, Fig. 3A, Fig. S2). Importantly, this illustrates that the introduction of a bifurcated modification into the fragment scaffold is tolerated and that fragment-binding affinity is maintained and not compromised by the introduction of simple linker groups. An *AfGna1-3* complex structure, which additionally contains Glc-6P, indicates that 3 is accessible to the fragment-binding site, despite narrow tunnels linking the fragment binding pocket to the enzyme active sites (Fig. 3B, Fig. S3). Like fragment hit 1 and 2, the important binding interactions and binding mode are maintained in the *AfGna1-3*-(Glc-6P) complex structure. In addition, the hydroxy functionality of the *N,N*-diethyl alcohol moiety displaces His-127-coordinating water molecules and interacts with the acceptor nitrogen atoms of the His-127 side chains (Fig. 3B). The introduction of a *para*-anilino anchoring point for bifurcation leverages the sp² character of the anilino-nitrogen, maintaining the planarity necessary at the branch point for correct *N,N*-diethyl alcohol orientation whereas *ortho*-dichloro substitution positions the fragment along the 2-fold crystallographic

axis (with a positional shift of 0.6 Å and 19° reorientation from fragment hit 1). As a result, bifurcation vectors are identically orientated toward both *Af*Gna1 active sites in contrast to the binding positions of fragment hit 1 and 2 (Fig. 3B). Moreover, the positioning of the Glc-6P 6.2 Å from the ethyl chain linker of 3, visible in the *Af*Gna1–3-(Glc-6P) complex structure (Fig. 3B, Fig. S3), is suggestive of fragment-linking approaches toward *Af*Gna1 inhibition, albeit challenging because of the limited size of the tunnel (46). In line with this, derivative 3 did not show inhibition against *Af*Gna1 in a recombinant enzyme assay, probably because of the inability of the *N,N*-diethyl alcohol moiety to extend into and block the enzyme active sites.

Globally, the structural differences between the co-complexes of *Af*Gna1 with 1, 2, and 3 are negligible (root mean square deviation ~0.35–0.40). However, when considering buried protein pockets and cavities in general, such seemingly trivial displacements may be of significance. Recent molecular dynamics simulations and experiments have suggested that large-scale side chain or secondary structure conformational changes are not a prerequisite for ligand binding to buried cavities but rather subtle and rapid movement of peptide structural elements can be sufficient to facilitate ligand ingress and egress (47–50). Although beyond the scope of this work, we speculate that the effects of any subtle movement of structural elements may propagate within *Af*Gna1 and be important for ligand-binding dynamics to the unusual buried dimer interface pocket. Taken together, we have discovered small molecules that target a fungal-specific binding hotspot at the Gna1 dimer interface, providing a scaffold for the development of Gna1 inhibitors based on these Gna1 binders.

Discussion

The current clinical management of invasive fungal infections is multifaceted, and as a consequence of limited antifungal pipeline progression, clinicians must preserve the finite number of agents available to manage infection (51). Recent decades witnessed an exodus of research and development into antifungal drug discovery by the pharmaceutical industry resulting in very few agents with novel mechanisms of action undergoing clinical trials. Clinical need prevails, despite optimal antifungal therapy, mortality rates remain around 50% although patient co-morbidities also contribute (1). In response, a multidisciplinary approach to develop new targets has been advocated (52).

Successful identification and validation of novel and ligandable targets is critical to the progression of the antifungal pipeline but the current shortage of unique targets combined with the intrinsic therapeutic hurdles introduced by the similarities of fungal and human enzymes only serves to complicate an already underappreciated and growing medical problem (1, 2, 53). An attractive antifungal target is the cell wall, a structure unique to fungi and essential for survival. The sugar polymers constituting the cell wall are all synthesized from sugar nucleotide precursors. Here, we used a multidisciplinary approach focused on an essential step in the biosynthesis of the sugar nucleotide UDP-GlcNAc, the precursor of cell wall chitin, as a target against *A. fumigatus*. Cell wall chitin has long been proposed as an antifungal target. The nikkomycins and polyoxins

are substrate analogues of UDP-GlcNAc and inhibit chitin synthesis *in vitro* with a predilection for *CHS-A* but fail *in vivo* primarily because of resistance in chitin synthase isoenzymes (54–56). Our approach circumvents this failure by targeting Gna1, a GNAT-family enzyme with no isoforms. Inhibitors targeting other GNAT family members are known in the literature (57, 58) with bisubstrate inhibitors, molecules incorporating tethered mimics of both substrate and cofactor, offering improved selectivity profiles compared with alternative approaches but harboring nonideal pharmacokinetic properties (59–63). However, such an approach is unlikely to offer robust selectivity in the case of Gna1 because of the high structural homology between fungal and human enzyme.

Previous failings in target-based drug discovery derive from an inadequate initial assessment of a molecular target in terms of physiological function (64). Although *GNA1* was shown to be essential for *in vitro* growth in *S. cerevisiae* and *C. albicans* (27, 29), intrinsic differences in gene function across fungi because of diverging biological and physiological functions, genetic redundancies, or scavenging pathways precludes predictions. Essentiality in yeasts does not automatically correlate in *A. fumigatus* and therefore this was experimentally validated to justify any potential investment in future inhibitor development. We demonstrated that genetic disruption of *A. fumigatus* *GNA1* results in a terminal phenotype under simulated physiological Glc and GlcNAc concentrations. This is further supported by our *in vivo* study using a neutropenic murine model of invasive aspergillosis with inhalation of *A. fumigatus* Δ *gna1* ~100-fold less pathogenic than control strains. However, contributions to the observed Δ *gna1* terminal phenotype from disruption to GPI anchor biosynthesis and *N*- and *O*-linked glycosylation cannot be excluded.

Although the finding corroborates previous work (29), we speculate that earlier *in vitro* studies were not sufficiently robust to detect physiologically relevant nutritional rescue strategies because if a gene were genuinely essential it would result in avirulence rather than attenuation. Gene deletion strategies incorporating nutritional rescue do not permit different levels of *GNA1* expression to be directly investigated. Altering the levels of extracellular GlcNAc reflects the ability of *A. fumigatus* to induce salvage pathways.

Synergy between cell wall active agents is increasingly seen as an attractive option for antifungal treatment (65). This is exemplified *in vitro* in *A. fumigatus* by the synergy between nikkomycin and the echinocandins (66). At present the echinocandins are advocated as salvage therapy in invasive aspergillosis in part because of their fungistatic activity (67). We show that *A. fumigatus* Δ *gna1* combined with caspofungin has a fungicidal effect on *in vitro* growth completely abolishing not only the terminal phenotype but also growth under maximal GlcNAc rescue. This provides further evidence that combining a β -glucan synthase inhibitor together with chitin inhibition is synergistic *in vitro*. A *GNA1* inhibitor could enhance the efficacy of the echinocandins and offer additional treatment modalities potentially as a resistance-sparing agent. In terms of biological importance, Gna1 is an attractive antifungal target.

In addition to our genetic validation of Gna1, we sought to identify compounds that would chemically phenocopy the bio-

logical phenotype. The inherent difficulty in leveraging inhibitor potency and selectivity between high-homology enzyme families among higher eukaryotic organisms is a challenging problem to fully resolve. This applies to Gna1 where deletion in *A. fumigatus* yields a terminal phenotype, but deletion of the murine orthologue leads to developmental phenotypes (68). We pursued a fragment-based approach to overcome this challenge by discovering a previously unidentified fungal selective pocket within AfGna1, positioned on the 2-fold axis generating the Gna1 biological homodimer. Crucially, this pocket is absent in the human orthologue, where it is occupied by larger side chains (arginines *versus* histidines in the fungal enzyme).

Our data show that fragment derivatives can recapitulate binding to this unusual fragment anchoring site, and rational design targeting this pocket allows the development of micromolar binders that do not target the human enzyme. Although not yet displaying AfGna1 inhibitory activity at high micromolar concentrations, the opportunities toward fungal-selective AfGna1 inhibition opened up by the discovery of fragments binding at this fungal-selective binding pocket is promising.

Moreover, with the exception of Arg-118, the amino acids that constitute the interface binding pocket are well-conserved between clinically relevant fungal Gna1 orthologues and HsGna1. Although sequence alignments have limited predictive capacity in determining the structural context of the interface pocket and its ligandability, this high degree of conservation warrants further structural and chemical exploration. This binding pocket may offer a ligandable handle against other fungal Gna1 orthologues, in particular, *C. auris*, which recently became the first fungal species listed as an urgent drug-resistant threat (11).

Identifying and pursuing novel avenues for inhibitor development to overcome organism selectivity barriers is critical to reversing the dearth in the antifungal pipeline, as is the timely identification and validation of unusual and novel targets, exemplified here by Gna1.

The work reported here also serves to further exemplify the ability of fragment-based screening to uncover unique and otherwise unforeseen avenues toward enzyme inhibition through the discovery of novel binding sites. Successful fragment campaigns that provide the foundation to potent inhibitors commonly begin with fragments that display no intrinsic enzymatic inhibitory activity (69). The fact that fragment hit 1 and derivatives (2 and 3) display a stable binding mode is often an important prerequisite to subsequent successful fragment development campaigns (70). Moreover, although small molecule therapeutics that bind to protein interfaces are uncommon, such approaches have been successfully utilized in structure-based design campaigns to inhibit enzyme targets. For example, HIV-1 and HIV-2 protease inhibitors, such as darunavir, bind to a cavity at the protein dimer interface located adjacent to the enzyme active site, forming contacts to the interface cavity and inhibiting enzymatic activity through hydrogen bonding to catalytically important D25 carboxylate side chains of both protease monomers thereby competitively inhibiting viral polypeptide access (71–73).

The AfGna1–3-(Glc-6P) complex structure offers a glimpse at potential avenues of inhibition via a combination of bifurca-

tion and fragment-linking approaches. Although a rare and challenging form of fragment development, successful fragment-linking campaigns ideally utilize the conjugation of fragment pairs with contrasting hydrophobic and polar ligand–protein–binding characteristics (46). Such characteristics are exemplified herein with fragment hit 1 and the sugar (or pseudosugar) substrate and therefore provide an incentive to further explore this approach. In parallel, exploring potential routes toward Glu-129 modulation of substrate binding has led to the synthesis of a submicromolar fragment binder and a key chemical scaffold which can now be further explored and modified to elicit the desired inhibitory response.

In summary, we have provided genetic validation of AfGna1 as an antifungal target and have discovered small molecules that target the Gna1 dimer interface, providing a platform for generating inhibitors of the enzyme. As demonstrated, our multidisciplinary approach, incorporating both genetic and chemical target assessment, is critical in determining target feasibility. Such approaches are not limited to fungal pathogens and are applicable to other infectious agents and are of particular relevance when considering targets with high sequence and structural homology to human enzymes.

Experimental procedures

Ethics statement

All animal experiments were performed by Evotec Ltd (UK) under UK Home Office License 40/3644 and ethically approved by The University of Manchester Standing Committee.

Strains, culture medium, and solutions

The genetic lineage and nutritional growth requirements of all strains used and generated in this work are provided in Table S1. Further details are provided in the supporting information.

Generation of *A. fumigatus* Δ gna1 mutant

To delete *GNA1*, construct p Δ gna1pyrG+ was designed to replace the entire 644-bp coding region of *GNA1* with the URA-blaster (also called the pyrG blaster) as a reusable selection marker by homologous recombination (74) (Fig. S1). Refer to the supporting information for all PCR primer sequences used in this work. The supporting information contains details of the experimental process including protoplast transformation, phenotypic screening, PCR screening, and Southern blot verification (Fig. S1) of *A. fumigatus* Δ gna1.

Generation of an *A. fumigatus* *GNA1*+6H::pyrG+ reconstituted strain

A construct p*GNA1*+6H was designed to replace the entire URA-blaster (8.3 kb) in the Δ gna1 mutant with the native *GNA1* sequence by homologous recombination (Fig. S1).

Experimental procedures culminating in Southern blot verification followed those for generation of the Δ gna1 mutant (see Table S1 and supporting information). To permit *in vivo* applications by restoring uridine and uracil autotrophy, the reconstituted strain (*GNA1*+6H::pyrG-) was complemented with pyrG by random integration. Primers were used to amplify a 1.9 kb portion of the *A. fumigatus* pyrG gene (75) from pXDRFP4 (76). Refer to supporting information for further details.

In vitro phenotypic analysis of *A. fumigatus* Δ *gna1* mutant

An agar dilution method was used to investigate *in vitro* growth in the presence of different concentrations of glucose and GlcNAc. Complete medium (CM) containing 90 mM, 150 μ M, and 1.5 μ M GlcNAc was supplemented with 0.1 mM glucose (Glc). CM supplemented with ≥ 50 mM Glc and 0 mM GlcNAc was a Δ *gna1* negative growth control. For preliminary evaluation of growth characteristics under the above conditions, suspensions containing serial dilutions of conidia (5 μ l, $1 \times 10^6 - 1 \times 10^3$) from the respective *A. fumigatus* strains were inoculated on solid medium and incubated at 37 °C for 48 h. To further assess the interaction between Glc and GlcNAc on growth, an agar microdilution method (77) was modified using flat-bottomed 96-well plates to generate a checkerboard assay. This enabled simultaneous testing of 285 different combinations of Glc and GlcNAc (final concentration range 0 mM to 50 mM, 2-fold serial dilutions) in a 200 μ l final volume (180 μ l of non-glucose CM agar and 10 μ l each of Glc and GlcNAc). Each well represented a different growth condition and was inoculated with *A. fumigatus* viable conidia (1×10^3) and incubated at 37 °C for 48 h. Wells were scored independently by two unblinded investigators using a numerical code (1, no growth/inoculum spotting to 7, growth equivalent to positive control) to obtain an average score per condition. Scores were converted to colors generating a growth heat map (Excel, Microsoft).

Germination was examined by inoculating liquid medium (20 ml, CM 110 mM Glc and CM 0.1 mM Glc + 90 mM GlcNAc) with conidia (1×10^5) and incubating in a Petri dish containing glass coverslips. At specified time points (5, 8, 12, and 24 h), the coverslips were removed, washed, and fixed (2 ml, 3.7% paraformaldehyde, 20 min). Coverslips were mounted onto glass slides and examined using a light microscope (Leica ICC50 HD, $\times 40$ objective lens magnification).

To examine the response of the *A. fumigatus* Δ *gna1* mutant to cell wall perturbing agents, suspensions of conidiophores (5 μ l, $1 \times 10^6 - 1 \times 10^2$) were inoculated onto solid 0.1 mM Glc CM + ≥ 50 mM GlcNAc in the presence of Calcofluor White, Congo Red, SDS (25–100 μ g/ml), and caspofungin (0.0625–0.5 μ g/ml). Following incubation (48 h), plates were examined. The experiment was performed three times. For analysis of cell wall architecture, conidia and mycelia were grown on solid medium, fixed, processed, and examined using transmission and scanning EM. Refer to [supporting information](#) for further details.

A. fumigatus Δ *gna1* infection models

Greater wax moth larvae, *G. mellonella* (Livefoods Direct, Sheffield, UK), were used as an infection model (36, 78, 79). Larvae were maintained at room temperature in wood shavings in the dark (80) and those in the sixth instar measuring 2.0 cm in length (~ 250 mg) were randomly assigned into groups. The *A. fumigatus* parental, Δ *gna1*, and reconstituted strains were inoculated into the hind proleg of larvae ($n = 30$ per group) as a 10 μ l PBS suspension of 5×10^5 resting conidia. Negative controls were inoculated with PBS only. Larvae were protected from light, incubated at 37 °C, and examined at 24 h intervals. Mortality, defined by lack of movement in response to stimuli

and discoloration (melanization) of the cuticle, was scored for 6 days.

An inhalational neutropenic murine model of invasive aspergillosis (37, 81) was performed by Evotec Ltd (UK) (see “Ethics statement”). As this was an initial proof of concept study and because of insufficient published literature using the parental strain in this particular murine model, group size by power analyses were unable to be performed for statistical differences between strains. Sharing the data generated in this study will inform future power calculations.

Male CD1 mice (Charles River, UK) weighing 23–26 g were used for all experiments and allowed to acclimatize for at least 7 days. Mice were randomly assigned to groups and housed in sterilized individually ventilated cages with HEPA filtered sterile air and sterile aspen chip bedding (changed every 3–4 days). Food and water were provided *ad libitum*. The room temperature was 22 °C (± 1 °C) with a relative humidity of 60% and maximum background noise of 56 dB. Mice were exposed to 12-h light/dark cycles.

Mice were rendered neutropenic with intraperitoneal cyclophosphamide (250 mg/kg on day -2 relative to infection, and 200 mg/kg on day $+3$) and subcutaneous cortisone acetate (200 mg/kg on day -2 and day $+3$). To generate conidial suspensions of the target inoculum (Table S2) for inhalational inoculation (infection) of mice, *A. fumigatus* parental, Δ *gna1*, and reconstituted strains were prepared according to a National Institutes of Health (NIH) standard operating procedure (82) with nutritional requirements as per Table S1. Viable cell counts were performed on serially diluted aliquots of each conidial suspension.

Each cage was randomly assigned to a study group and infected by an inhalation chamber system (37). On the day of infection, mice ($n = 4$ per group for experiment 1; $n = 6$ per group for experiment 2) were exposed for 1 h to 12 ml of PBS containing *A. fumigatus* conidia aerosolized via a Micro Mist[®] nebulizer (Teleflex Medical, Kernen, Germany) in an acrylic chamber. Enrofloxacin (50 ppm, 0.2 ml/water bag) was administered from day -3 to day $+10$ to prevent bacterial infection. Mice were weighed and monitored at least daily and the study refined using surrogate markers for invasive pulmonary aspergillosis. Any mouse with signs of illness and distress (see next paragraph) were culled immediately (terminal anesthesia or cervical dislocation) and death recorded as being the next day.

A scoring system was used based on the following parameters: weight loss; hypothermia less than 32 °C; appearance (hunching, poor coat condition); clinical signs: respiration (noisy labored breathing); tremors/convulsions; unprovoked behavior (little peer interaction); and provoked behavior (subdued when stimulated).

Statistical analyses

Biological experiments were performed at least three independent times with the exception of EM. Statistical analyses were carried out using GraphPad Prism (v5.0b). Data are expressed as the mean \pm S.D. (Fig. 1C). To compare more than two groups, one-way ANOVA test with Bonferroni's multiple comparison test was used (Fig. 1C). For comparison of survival

curves, the Kaplan-Meier Log-Rank (Mantel-Cox) Test was performed (Fig. 2A only).

AfGna1 protein expression and purification

AfGna1 was purified as described earlier (39). Briefly, the sequence encoding the protein was inserted into a pGEX6P-1 vector for expression of a GST tagged fusion that contains PreScission protease cleavage site. For protein expression, the plasmid was transformed into *Escherichia coli* BL21(DE3) pLysS. Cell cultures were grown to an A_{600} of 0.8 and expression was initiated by the addition of 250 μ M isopropyl 1-thio- β -D-galactopyranoside at 17 °C. The cultures were then allowed to grow for a further 16 h before harvesting by centrifugation. After lysis, affinity chromatography and GST tag cleavage, eluted protein was concentrated and further purified by size exclusion chromatography using a Superdex 75, 26/60 column. Corresponding fractions confirmed by SDS-PAGE were pooled concentrated, flash frozen in liquid nitrogen, and stored at –80 °C.

AfGna1 fragment screen and binding affinity measurements

To identify a potential fragment binder of AfGna1, the fragment library of the University of Dundee Drug Discovery Unit (DDU) comprising of 652 structurally diverse compounds was screened using BLI (41–43). BLI experiments were performed on an Octet Red 384 system (Forte Bio, U.S.A.) using super streptavidin (SSA) biosensors and biotinylated-AfGna1. Primary hits identified from initial screening were defined as fragments with a response rate greater than three robust standard deviation units above the median. Confirmation of fragment hit binding was performed with a follow-up concentration dose-response series. Binding isotherms were fitted and visualized using Octet software (Forte Bio, U.S.A.) to derive dissociation constants (K_d). Refer to [supporting information](#) for further experimental details.

AfGna1 crystallography

AfGna1 crystallization was carried out as described earlier (39). Crystallization was manually performed using a sitting drop approach by combining protein solution previously incubated with AcCoA for 2 h on ice with reservoir solution in a 1:1 ratio and equilibrated against 60 μ l reservoir solution. AfGna1 crystals resembling bipyramidal or bar shape were generally observed after 72 h incubation at 20 °C.

For fragment soaking, a soaking solution for each confirmed fragment hit was prepared by combining a fragment DMSO stock solution (50–200 mM) with crystallization reservoir solution. After incubation (1 h to overnight) at 20 °C, crystals were transferred to a cryoprotectant using a nylon loop, immersed for 10 s and flash frozen in liquid nitrogen.

Diffraction data were collected at the European Synchrotron Radiation Facility (ESRF), Grenoble (France) and the Diamond Light Source, Oxford (UK). The diffraction data were automatically processed and scaled by *xia2* (83). An initial AfGna1-1 complex was solved by molecular replacement using *MOLREP* (84) using AfGna1 apo structure (PDB ID 2VEZ (90)) as a search molecule. Refinement was done using *REFMAC5* (85) and model building with *Coot* (86). Well-defined electron density

for fragment hit 1 was evident at the dimer interface. An apo version of this new model served as a search molecule for the determination of subsequent complexes by molecular replacement. Where appropriate, ARP/wARP (87) was used to partially build the model before further refinement. PRODRG (88) was used to generate coordinates and topologies of the compounds (ligands). Ligands were not included until their conformations were completely defined by unbiased σ A-weighted $F_o - F_c$, φ_{calc} electron density maps.

Chemical synthesis of fragment derivatives

2-chloro-3-(1,2,4-triazol-3-yl)aniline, compound 2—3-amino-2-benzoic acid (0.5 g, 2.90 mmol) was dissolved in dichloromethane/*N,N*-dimethylformamide (7 ml:2 ml). To the solution was added *N,N*-diisopropylethylamine (1.5 ml, 8.70 mmol), 1-hydroxybenzotriazole hydrate (470 mg, 3.48 mmol), and *N*-(3-dimethylaminopropyl)-*N*-ethylcarbodiimide hydrochloride (667 mg, 3.48 mmol). The solution was stirred at room temperature under argon over 20 min, ensuring that all solid material had dissolved. Ammonium hydroxide solution (25%, 0.35 ml) was added dropwise to the stirring solution, forming a pale yellow suspension which was then left overnight to stir. After aqueous workup (~5–10 ml ddH₂O), extraction with ethyl acetate (4 × 15 ml), and drying over sodium sulfate, the crude material (~440 mg) was concentrated and rapidly purified over a silica plug (ethyl acetate → ethyl acetate/MeOH). Eluted fractions were analyzed by LC-MS, confirming product formation, and fractions containing the desired intermediate amide product were combined (320 mg, 64% yield). Without further purification, the intermediate amide (314 mg, 1.84 mmol) was dissolved in *N,N*-dimethylformamide dimethyl acetal (6 ml) and was refluxed, under argon, at 115 °C for 15 h. After refluxing, the solution was cooled and evaporated to dryness *in vacuo*. In a separate receptacle, 50–60% hydrazine hydrate solution (150 μ l) was added slowly (dropwise) to acetic acid (9 ml). This solution was then rapidly transferred to the dried intermediate and refluxed at 95 °C for 7 h. After reflux, the solution was cooled and evaporated to dryness *in vacuo*. The crude material was dry loaded onto silica and was directly purified by Flash Chromatography (heptane/ethyl acetate → ethyl acetate → ethyl acetate/MeOH). Eluted fractions were analyzed by LC-MS, appropriately combined, and evaporated to dryness and resulted in an off-white solid (post-lyophilization from 50% acetonitrile and ddH₂O) (52 mg, 14% yield, R_f 0.76, ethyl acetate). Overall, 9% yield over two steps: ¹H NMR (500 MHz, MeOH-D₄): δ = 8.57 (s, 1 H), 7.21 (dd, *J* = 7.6 Hz, 7.6 Hz, 1 H), 7.06 (dd, *J* = 8.1 Hz, 1.6 Hz, 1H), 7.01 (dd, *J* = 7.5 Hz, 1.6 Hz, 1H). ¹³C NMR (101 MHz, MeOH-D₄): δ = 156.64, 147.33, 145.86, 128.61, 125.90, 121.43, 119.22, 118.60. HRMS-TOF: Theo. *m/z* 195.0432; obs. *m/z* 195.0428; [M+H], C₈H₈ClN₄.

2,2'-(3,5-dichloro-4-(1,2,4-triazol-3-yl)phenyl)azanediyl-bis(ethan-1-ol), compound 3—2,6-dichloro-4-nitrobenzamide (1 g, 4.25 mmol) was dissolved in ethanol (12 ml) followed by the addition of tin (II) chloride dihydrate (4.8 mg, 21.30 mmol). The resulting solution was refluxed at 70 °C overnight at which point the solution was cooled, poured into ice water (50 ml), and pH adjusted (to pH ~8–9) using 4N aqueous sodium hydroxide, where it was allowed to stir for 30 min. The resulting

solid was removed by filtering over celite and the aqueous filtrate was then extensively extracted with ethyl acetate (6 × 25 ml). The organic phases were combined, dried over sodium sulfate, and evaporated to dryness *in vacuo*, resulting in an off-white solid (intermediate aniline) which was used without further purification (72% yield).

The intermediate aniline (250 mg, 1.22 mmol) was dissolved in acetic acid (2.5 ml), followed by the addition of ~3 M ethylene oxide–THF solution (2.5 ml). The resulting solution was stirred at room temperature for 24 h at which point the solvent was evaporated to near dryness *in vacuo*. The crude material was dry loaded onto silica and was directly purified by Flash Chromatography (ethyl acetate → ethyl acetate/MeOH). Eluted fractions were analyzed by LC-MS, appropriately combined and evaporated to dryness, resulting in a mixture of unreacted starting material, *N*- and *N,N*-ethyl alcohol products. The reaction and subsequent FLASH purification were then repeated using the combined unreacted starting material and monofunctionalized product to improve the conversion to the *N,N*-ethyl alcohol product (145 mg, 40% yield).

The *N,N*-ethyl alcohol product (100 mg, 0.34 mmol) was dissolved in a 1:1 mixture of pyridine and acetic anhydride (10 ml) and was left to stir overnight, at which point the solution was evaporated to dryness followed by trituration of the resulting oil with toluene (2 × 5 ml) (quantitative hydroxyl group protection). Without further purification, the *O*-acetyl protected material (~125 mg, ~0.34 mmol) was dissolved in *N,N*-dimethylformamide dimethyl acetal (5 ml) and was refluxed, under argon, at 115 °C for 24 h. After refluxing, the solution was cooled and evaporated to dryness *in vacuo*. In a separate receptacle, 50–60% hydrazine hydrate solution (8 eq, 85 μl) was added slowly (dropwise) to acetic acid (8 ml). This solution was then rapidly transferred to the dried intermediate and was refluxed at 95 °C for 15 h. After reflux, the solution was cooled and evaporated to dryness *in vacuo*. The crude material was dry loaded onto silica and was directly purified by Flash Chromatography (heptane/ethyl acetate → ethyl acetate). Eluted fractions were analyzed by LC-MS and appropriately combined and evaporated to dryness, resulting in a white foam (85 mg, 48% yield). The resulting 1,2,4-triazole (40 mg, 0.10 mmol) was then *O*-acetyl deprotected using a 1:1 solution of 1,4-dioxane and 1.5 M aqueous sodium hydroxide (pH ~10). The reaction was monitored by TLC and on completion, the basic solution was neutralized using Amberlite IR-120 H⁺ ion-exchange resin. The resin was rinsed and the resulting solution was lyophilized. The lyophilized material was then purified by Flash Chromatography (2 g silica; gradient elution, 1:1 ethyl acetate/heptane → ethyl acetate → 3:1 ethyl acetate/methanol). Eluted fractions were analyzed by LC-MS and appropriately combined and evaporated to dryness (20 mg, 63% yield; R_f 0.41, ethyl acetate), with an overall yield of 9% yield over five steps: ¹H NMR (500 MHz, MeOH-D₄): δ = 8.34 (br, s, 1 H), 6.88 (s, 2 H), 3.76 (t, J = 6.0 Hz, 4 H), 3.58 (t, J = 6.0 Hz, 4 H). ¹³C NMR (101 MHz, MeOH-D₄): δ = 153.21–151.14 (br, s), 144.66, 137.53, 133.52, 112.14, 59.95, 54.58. HRMS-TOF: Theo. *m/z* 317.0567; obs. *m/z* 317.0573; [M+H], C₁₂H₁₅Cl₂N₄O₂.

Data availability

The structures presented in this paper have been deposited in the Protein Data Bank with accession codes 6TDH, 6TDG, and 6TDF. All remaining data are included within the manuscript and [supplementary information](#) or can be shared upon request (Prof. Daan M. F. van Aalten, University of Dundee, dmfvanaalten@dundee.ac.uk).

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Abbreviations—The abbreviations used are: GPI, glycosylphosphatidylinositol; Glc-6P, glucose-6-phosphate; SEM, scanning electron microscopy; TEM, transmission electron microscopy; AcCoA, acetyl coenzyme A; *Hs*, *Homo sapiens*; BLI, bio-layer interferometry; CM, complete medium.

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